

ENDOPARASITES OF WILD BIRDS FROM CAMPUS AREA AND ZOOBOTANICAL PARK, AT THE FEDERAL UNIVERSITY OF ACRE, RIO BRANCO - ACRE

Angela Silva de Almeida Brito¹
Edson Guilherme²
Francisco Glauco de Araújo Santos²
Rosiane Portela de Mesquita¹
Fábio Augusto Gomes²

BRITO, A. S. de A.; GUILHERME, E.; SANTOS, F. G. de A.; MESQUITA, R. P. de; GOMES, F. A. Endoparasites of wild birds from Campus area and Zoobotanical Park, at the Federal University of Acre, Rio Branco –Acre. *Arq. Ciênc. Vet. Zool. UNIPAR*, Umuarama, v. 20, n. 3, p. 117-122, jul./set. 2017.

ABSTRACT: This study was developed at the Campus (C) and the Zoo-botanical Park (PZ) at the Federal University of Acre, with the purpose of identifying endoparasites in the avifauna of those locations. The birds were captured using 10 mist nets. The nets were set from 1 p.m. to 5 p.m.. The faecal samples collected were stored in Merthiolate, Iodine and Formaldehyde (MIF). Willis and direct methods were used for the diagnostic of endoparasites. Two hundred and eighty-seven birds were captured, with 34 recaptures, which resulted in 253 different specimens – 158 on the campus and 95 at the PZ. The birds caught were from 58 different species, belonging to 25 families and 12 orders. Fecal samples from 193 birds belonging to 49 species from 11 orders and 18 families were obtained and analyzed. The following endoparasites were identified: *Eimeria* sp., *Entamoeba coli*, *Entamoeba histolytica*, *Iodamoeba bütschlii*, *Isospora* sp., *Enterobius vermicularis*, *Heterakis gallinarum*, *Ascaridia galli*, *Strongyloides avium*, *Capillaria* spp., *Ancylostoma* spp., *Raillietina* sp., *Choanotaenia* sp., *Microphallus* sp., and *Echinostoma revolutum*. *Eimeria* sp. was the most frequent parasite found in most bird species. The results showed a high level of endoparasitic infection in the aviary community in the two study areas.

KEYWORDS: Acre. Amazon. Avifauna. *Eimeria*.

ENDOPARÁSITOS EN AVES SILVESTRES DEL ÁREA DEL CAMPUS Y DEL PARQUE ZOOBOTÁNICO DE LA UNIVERSIDAD FEDERAL DEL ACRE, RIO BRANCO - ACRE

RESUMEN: El estudio se realizó en el Campus (C) y en el Parque Zoobotánico (PZ), de la Universidad Federal de Acre, con el objetivo de identificar los endoparásitos presentes en la avifauna de estos locales. Las aves fueron capturadas utilizando 10 redes de niebla. Las redes fueron armadas en el período de las 13h a las 17h. Las muestras fecales obtenidas se conservaron en Merthiolate, Yodo y Formaldehído (MIF). Para el diagnóstico de los endoparásitos se utilizaron los métodos directo y de Willis. Se capturaron 287 aves, habiendo ocurrido 34 recapturas, lo que resultó en 253 especímenes distintos, siendo 158 en el Campus y 95 en el PZ. Las aves capturadas pertenecían a 58 especies, distribuidas en 25 familias y 12 órdenes. Se obtuvieron y analizaron las muestras fecales de 193 aves, pertenecientes a 49 especies de 11 órdenes y 18 familias. Los endoparásitos identificados: *Eimeria* sp., *Entamoeba coli*, *Entamoeba histolytica*, *Iodamoeba butschlii*, *Isospora* sp., *Enterobius vermicularis*, *Heterakis gallinarum*, *Ascaridia galli*, *Strongyloides avium*, *Capillaria* sp., *Ancylostoma* sp., *Raillietina* sp., *Choanotaenia* sp., *Microphallus* sp. y *Echinostoma revutum*. La *Eimeria* sp. fue el parásito que estuvo presente en mayor número de especies de aves. Los resultados evidenciaron un alto nivel de infección de endoparásitos en la comunidad de aves, presentes en las dos áreas del estudio.

PALABRAS CLAVE: Acre. Amazonia. Avifauna. *Eimeria*.

ENDOPARASITAS DE AVES SILVESTRES DA ÁREA DO CAMPUS E DO PARQUE ZOOBOTÂNICO, DA UNIVERSIDADE FEDERAL DO ACRE, RIO BRANCO - ACRE

RESUMO: O estudo foi realizado no Campus (C) e no Parque Zoobotânico (PZ), da Universidade Federal do Acre, com o objetivo de identificar os endoparasitas presentes na avifauna destes locais. As aves foram capturadas utilizando-se 10 redes de neblina. As redes foram armadas no período das 13h às 17h. As amostras fecais obtidas foram conservadas em Merthiolate, Iodo e Formaldeído(MIF). Para diagnóstico dos endoparasitas foram utilizados os métodos direto e de Willis. Foram capturadas 287 aves, tendo ocorrido 34 recapturas, o que resultou em 253 espécimes distintas, sendo 158 no Campus e 95 no PZ. As aves capturadas pertenciam a 58 espécies, distribuídas em 25 famílias e 12 ordens. Foram obtidas e analisadas as amostras fecais de 193 aves, pertencentes a 49 espécies de 11 ordens e 18 famílias. Os endoparasitas identificados: *Eimeria* sp., *Entamoeba coli*, *Entamoeba histolytica*, *Iodamoeba butschlii*, *Isospora* sp., *Enterobius vermicularis*, *Heterakis gallinarum*,

DOI: 10.25110/arqvet.v20i3.2017.6363

¹Mestre – Pós-Graduação em Ciências, Inovação e Tecnologia para Amazônia – Universidade Federal do Acre, UFAC, Rio Branco, AC, Brasil, E-mail: angelabrito16@gmail.com; rosiiportela@gmail.com

²Professor Doutor - Centro de Ciências Biológicas e da Natureza – Universidade Federal do Acre, UFAC, Rio Branco, AC, Brasil, E-mail: guilherme@ufac.br; edson@uol.com.br; fcoglauco@ufac.br e agosto.ufac@gmail.com

Ascaridia galli, *Strongyloides avium*, *Capillaria* sp., *Ancylostoma* sp., *Raillietina* sp., *Choanotaenia* sp., *Microphallus* sp. e *Echinostoma revolutum*. A *Eimeria* sp. foi o parasito que esteve presente em maior número de espécies de aves. Os resultados evidenciaram um alto nível de infecção de endoparasitas na comunidade de aves, presentes nas duas áreas de estudo.

PALAVRAS-CHAVE: Acre. Amazônia. Avifauna. *Eimeria*.

Introduction

The birds are a group of vertebrates found in virtually all regions of the planet. Birds are among the most easily-recognized of animals, due to their ample geographic distribution, and the predominantly diurnal habits of most species (SICK, 1985).

Wild birds are subject to a variety of health problems, which may affect their survival and reproduction. Parasitic infestation is one of the most prominent of these problems, not only because of the frequency of occurrence, but also due to their potential impact, provoking serious infections and even death in a large proportion of the bird populations found in intensely parasitized regions (REED et al., 2003; MARIETTO et al., 2009).

Parasitism has been reported in birds by a number of authors (FREITAS et al., 2002; LUZ et al., 2005; JACOBSEN et al., 2006; MARIETTO et al., 2009). The most common avian endoparasites are protozoa, nematodes, cestodes, and trematodes (REY, 2008).

Eimeria and *Isospora* are among the most prominent protozoan parasites of birds, both domestic and wild species, causing persistent illness, especially in captivity (MARIETTO et al., 2009).

Eimeria causes coccidiosis, a common infectious disease in birds, usually limited to the digestive tract. Infection occurs through ingestion of oocysts found in the nest, soil, food or contaminated water. After ingestion, the protozoa through a series of intracellular and extracellular stages, asexual and sexual, to produce viable oocysts which are excreted in the feces (ROSE, 1987).

Nematodes are considered to be the most pathogenic endoparasites, and may have a considerable economic impact on the poultry industry. The occurrence of infections by these endoparasites is more common in free-range domestic birds and in the wild, but it is relatively uncommon in confined poultry (TAYLOR et al., 2010). These parasites may be transmitted horizontally, directly between birds, through the ingestion of larvae, or by the indirect cycle, which requires an intermediate host, such as a mollusc or an insect. Nematodes include *Syngamus trachea*, *Dispharynx*, *Tetramers*, *Ascaridia*, and *Capillaria* (TAYLOR et al., 2010).

While they are less pathogenic than nematodes, cestodes can also cause disease in birds, in particular, severe hemorrhagic enteritis. This endoparasite is also important because of its ability to infect a large number of individuals in a population. Over a thousand species of cestodes have been identified in domestic and wild birds, such as *Davineaproglottina* and *Raillietina tetragona* (TAYLOR et al., 2010).

The trematodes that infect the bird intestine include *Echinostoma revolutum*, which can cause diarrhea, malnutrition and fatigue. A high parasite load of this worm can lead to death through the perforation of the intestine of the infected individual (TAYLOR et al., 2010).

The identification of endoparasites is essential for the understanding of the prevalent species that infect a wild

bird fauna, in addition to the development and implementation of effective measures of control. In this context, the present study aimed to identify the endofauna of wild birds captured on the campus and the adjacent Zoobotanical Park, which are part of the Federal University of Acre in Rio Branco, in the western Brazilian Amazon basin.

Materials and Methods

Birds were captured for the collection of fecal samples in two areas of the Federal University of Acre (UFAC), the Campus (C) proper, and the adjacent Zoobotanical Park (ZP). The UFAC campus is located in the urban zone of Rio Branco, the capital of the Brazilian state of Acre, which is located 7 km from the city center, on the BR-364 federal highway.

Birds were captured between September 2014 and March 2015. Ten mist-nets (12 m x 2 m x 36 mm) were used, and were installed close to the ground, being set for four hours per day (13:00–17:00 h) on two days a week at each collection site. The nets were inspected regularly, according to the frequency of captures.

The captured birds were marked with numbered metal rings provided by CEMAVE/IBAMA. The bird species were identified by *Dr. Edson Guilherme* and the biologist *Rosiane Portela*, both of the UFAC Ornithology Laboratory. The scientific nomenclature of the Brazilian Committee of Ornithological Records (CBRO, 2014) was applied for the identification of the species.

Once identified and banded, the birds were placed in cloth bags containing absorbent paper until they defecated or up to a maximum of 40 minutes. The birds were then released, even if they had not defecated by the end of the 40-minute period. The fecal samples obtained from each bird were packed separately in plastic pots containing a preservative solution of Merthiolate-Iodine-Formaldehyde (MIF) for subsequent parasitological examination. The presence of endoparasites was diagnosed using the direct and Willis-Mollay methods, as described by De Carli (2001), using 10x and 40x lenses for the visualization and identification of the parasitic forms. This study was approved by the UFAC Ethics Committee on the Use of Animals (CEUA-UFAC), and the collection of wild animals was authorised by SISBIO 232691.

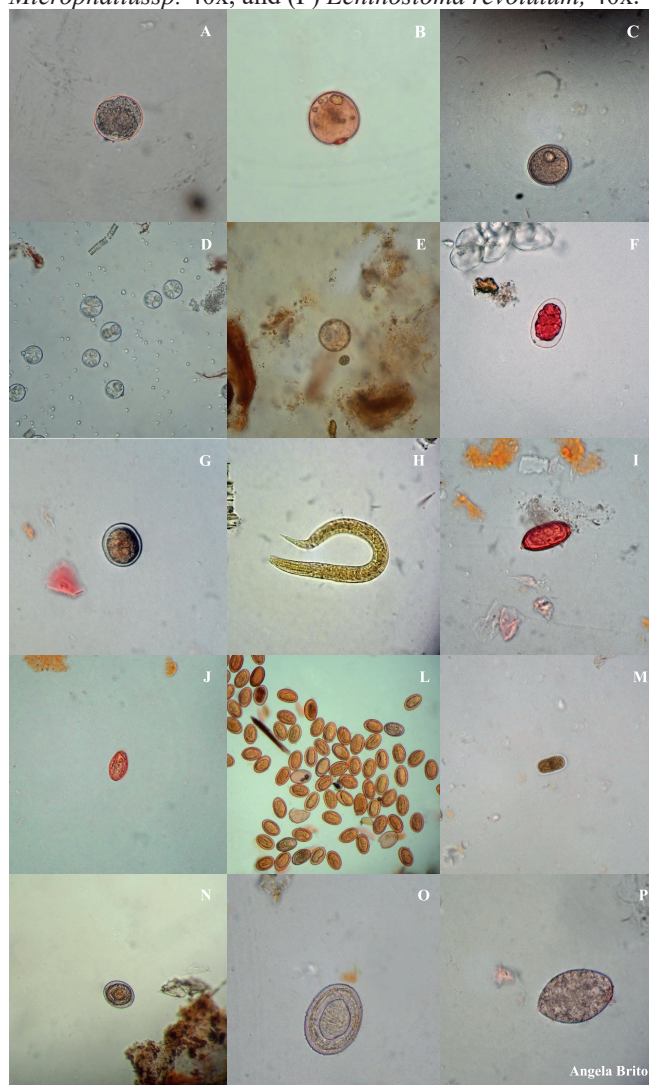
Results and Discussion

During the study, a total of 287 birds were captured, representing 253 different individuals and 34 recaptures. Of this total, 158 individuals were captured on the Campus, and 95 in the Zoobotanical Park. The captured birds belonged to 58 species, representing 25 families and 12 orders. In a previous study at UFAC between July 1998 and February 2000, Guilherme (2001) recorded 150 bird species in 36 families, using seven mist nets and complementary observations with binoculars.

Fecal samples were obtained from 193 birds (130

from the Campus and 63 from the Zoobotanical Park), belonging to 49 species in 18 families and 11 orders. A total of 15 endoparasite species were recorded in the parasitological analyses: *Eimeria* sp., *Entamoeba coli*, *Entamoeba histolytica*, *Isospora* sp., *Iodamoeba butschlii*, *Ancylostoma* sp., *Ascaridia galli*, *Strongyloides avium*, *Capillaria* sp., *Enterobius vermicularis*, *Heterakis gallinarum*, *Raillietina* sp., *Choanotaenia* sp., *Microphallus* sp., and *Echinostoma revolutum*. These taxa were identified based on the observation of oocysts, eggs and larvae under light microscopy (Figure 1).

Figure 1. Endoparasites present in the fecal samples analyzed. (A) *Eimeria* sp., 40x, (B) *Entamoeba coli*, 40x, (C) *Entamoeba histolytica*, 40x, (D) *Isospora* sp., 10x, (E) *Iodamoeba butschlii*, 40x, (F) *Ancylostoma* sp., 40x, (G) *Ascaridia galli*, 40x, (H) *Strongyloides avium*, 40x, (I) *Capillaria* sp., 40x, (J) *Enterobius vermicularis*, 40x, (L) *Heterakis gallinarum*, 40x, (M) *Raillietina* sp., 40x, (N) *Choanotaenia* sp., 40x, (O) *Microphallus* sp., 40x, and (P) *Echinostoma revolutum*, 40x.



Eimeria sp. and *Isospora* sp. were the most generalist endoparasite species, being recorded in 23 and 22 bird species, respectively (Table 1). The highest *Eimeria* sp. infection rate was recorded in *Jacana jacana*, with 16 of the 20 (80%) birds captured being infected with this parasite. *Eimeria* sp. was also found in the fecal samples of *Chloro-*

eryle americana, *Cacicus cela*, *Sturnella militaris*, *Progne chalybea*, *Sporophila castaneiventris*, *Colaptes punctigula* and *Phaeomyias murina*, *Tringa solitaria*, *Columbina talpacoti*, *Leptopogon amaurocephalus*, *Brotogeris sanctithomae*, *Thamnophilus schistaceus*, *Troglodytes musculus*, *Ramphocelus carbo*, *Crotophaga ani*, *Myiozetetes similis*, *Turdus ignobilis*, *Momotus momota*, *Ammodramus aurifrons*, *Turdus hauxwelli*, *Myiozetetes cayanensis*, and *Tyrannus melancholicus*. In a study of the gastrointestinal parasites of pigeons captured in urban areas of Lages, Santa Catarina (Brazil), Marques et al. (2007) found that 86.05% (37 of 43) of the birds were parasitized by protozoans, with *Eimeria* sp. oocysts being detected in all cases.

The *Isospora* sp. infection rate was highest in *Turdus ignobilis*, with 12 of the 20 (60%) samples analyzed being infected. *Isospora* sp. was also present in *Dendrocygna viduata*, *Vanellus chilensis*, *Hemitriccus flammulatus*, *Cacicus cela*, *Catharus swainsoni*, *Cantorchilus leucotis*, *Pheugopedius genibarbis*, *Troglodytes musculus*, *Tangara episcopus*, *Turdus ignobilis*, *Turdus hauxwelli*, *Rhynchocyclus olivaceus*, *Sporophila castaneiventris* and *Thamnophilus schistaceus*, *Ramphocelus carbo*, *Tringa solitaria*, *Machaeropterus pyrocephalus*, *Crotophaga ani*, *Tyrannus melancholicus*, *Columbina talpacoti*, *Pipra fasciicauda*, and *Jacana*.

Eimeria sp. and *Isospora* sp. are among the endoparasites most commonly found in birds. In a study of wild birds, Marietto et al. (2009) reported that species of five of the 12 orders analyzed were infected. The coccidians (*Eimeria* and *Isospora*) were the most common parasites, being found principally in individuals of the order Passeriformes.

As the causal agent of coccidiosis, *Eimeria* sp. is considered to be the most deleterious of the protozoans that cause disease in birds, and is a major problem for the poultry industry. The symptoms caused by this parasite include enteritis and diarrhea, and synergic effects with other diseases can make the infection even more severe (ALLEN; FETTERER, 2002).

In the present study, *Capillaria* sp. was recorded in the birds *Dendrocincla fuliginosa*, *Cacicus cela*, *Sporophila castaneiventris*, *Momotus momota*, *Jacana jacana*, *Myiozetetes similis*, *Turdus ignobilis* and *Tyrannus melancholicus*. *Capillaria* is potentially one of the most pathogenic helminths and may have grave effects on confined poultry. Mapeli et al. (2003) observed natural helminth infections, including *Capillaria* sp., in partridges being bred in captivity in the municipality of Jaboticabal, São Paulo (Brazil).

Other helminths recorded in the present study included *Ascaridia galli*, *Heterakis gallinarum* and *Raillietina* sp., which were also observed by Giovannoni and Kubiak (2001) during autopsies of domestic animals. These authors recorded *A. galli*, *H. gallinarum* and *Raillietina boniniem* in different portions of the digestive tract of *Columba livia domestica*.

Strongyloides avium was found in nine bird species at UFAC, *Cacicus cela*, *Sporophila castaneiventris*, *Momotus momota*, *Crotophaga ani*, *Jacana jacana*, *Turdus ignobilis*, *Myiozetetes similis*, *Myiozetetes cayanensis*, and *Pipra fasciicauda*. Between August 2010 and July 2012, Snak et al. (2014) inventoried 228 fecal samples of 37 bird species in 22 enclosures of Danilo Galafassi Municipal Park in Cascavel, Paraná (Brazil), and recorded five parasite genera,

Strongyloides, *Eimeria*, *Capillaria*, *Deletrocephalus*, and *Isoospora*.

Here, *Entamoeba histolytica* was found in the birds *Sporophila castaneiventris*, *Aratinga weddellii*, of *Tringa solitaria*, *Ammodramus aurifrons*, *Jacana jacana*, *Ramphocelus carbo*, *Columbina talpacoti*, *Crotophaga ani*, and *Tyrannus melancholicus*. *Entamoeba coli* was present in the birds *Dendrocincla fuliginosa*, *Ammodramus aurifrons*, *Columbina talpacoti*, *Crotophaga ani*, and *Jacana jacana*. These endoparasites were also reported by Marietto et al. (2009), who recorded *Entamoeba* sp. in the fecal samples of wild birds held in captivity, which were treated at the Ornithopathology and Parasitic Diseases laboratories at the Veterinary Hospital of the College of Veterinary Medicine and Animal Science at Paulista State University in São Paulo (Brazil).

In the present study, *Ancylostoma* sp. occurred in the feces of 10 bird species, *Geotrygon montana*, *Jacana jacana*, *Pitangus sulphuratus*, *Tringa solitaria*, *Hypocnemis subflava*, *Turdus hauxwelli*, *Turdus ignobilis*, *Crotophaga ani*, *Columbina talpacoti*, and *Myiozetetes similis*. *Enterobius vermicularis* was found in five birds, *Sturnella militaris*, *Leptopogon amaurocephalus*, *Tringa solitaria*, *Columbina talpacoti* and *Jacana jacana*. *Microphallus* sp. occurred in seven birds, *Tringa solitaria*, *Jacana jacana*, *Myiozetetes*

cayanensis, *Tyrannus melancholicus*, *Myiozetetes similis*, *Columbina talpacoti* and *Turdus ignobilis*. *Choanotaenia* sp. was found in six birds, *Aratinga weddellii*, in *Tringa solitaria*, *Sturnella militaris*, *Tangara episcopus*, *Troglodytes musculus*, and *Turdus ignobilis*.

Echinostoma revolutum was present in 11 birds, including all the fecal samples examined from the species *Chloroceryle inda*, *Pheugopedius genibarbis*, and *Pitangus sulphuratus*. It was also found in the feces of *Tringa solitaria*, *Leptopogon amaurocephalus*, *Momotus momota*, *Sturnella militaris*, *Crotophaga ani*, *Turdus hauxwelli*, *Jacana jacana* and *Turdus ignobilis*. *Iodamoeba butschlii* was recorded only in *Brotogeris sanctithomae*.

Freitas et al. (2002) analyzed wild birds kept in captivity in the Brazilian state of Pernambuco and reported a high incidence of parasites. In this study, 320 of the 685 birds analyzed were parasitized with *Capillaria* sp., *Strongyloides* sp., *Ascaridia* sp., *Heterakis* sp., Spiruroidea, Cestodea, Trematoda, Coccidia, *Entamoeba coli*, *E. histolytica* and *Balantidium coli*. In addition to these endoparasites, four other taxa were recorded in the present study, *Entamoeba coli*, *Iodamoeba butschlii*, *Choanotaenia* sp. and *Echinostoma revolutum*, reinforcing the diversity of bird endoparasites found in the study region.

Table 1. Endoparasites identified in the present study, and the bird species in which they were recorded, based on the analysis of fecal samples collected at the Federal University of Acre, in Rio Branco, Brazil.

Parasite species	Bird species infected
<i>Ancylostoma</i> sp.	<i>Jacana jacana</i> , <i>Tringa solitaria</i> , <i>Pitangus sulphuratus</i> , <i>Turdus ignobilis</i> , <i>Myiozetetes similis</i> , <i>Columbina talpacoti</i> , <i>Geotrygon Montana</i> , <i>Crotophaga ani</i> , <i>Hypocnemis subflava</i> , <i>Turdus hauxwelli</i> .
<i>Ascaridia galli</i>	<i>Vanellus chilensis</i> , <i>Jacana jacana</i> , <i>Columbina talpacoti</i> , <i>Crotophaga ani</i> , <i>Hemitriccus flammulatus</i> .
<i>Capillaria</i> sp.	<i>Jacana jacana</i> , <i>Momotus momota</i> , <i>Dendrocincla fuliginosa</i> , <i>Cacicus cela</i> , <i>Sporophila castaneiventris</i> , <i>Turdus ignobilis</i> , <i>Myiozetetes similis</i> , <i>Tyrannus melancholicus</i> .
<i>Choanotaenia</i> sp.	<i>Tringa solitaria</i> , <i>Sturnella militaris</i> , <i>Tangara episcopus</i> , <i>Aratinga weddellii</i> , <i>Turdus ignobilis</i> , <i>Troglodytes musculus</i> .
<i>Eimeria</i> sp.	<i>Jacana jacana</i> , <i>Tringa solitaria</i> , <i>Columbina talpacoti</i> , <i>Chloroceryle americana</i> , <i>Momotus momota</i> , <i>Crotophaga ani</i> , <i>Cacicus cela</i> , <i>Sturnella militaris</i> , <i>Progne chalybea</i> , <i>Ammodramus aurifrons</i> , <i>Leptopogon amaurocephalus</i> , <i>Ramphocelus carbo</i> , <i>Sporophila castaneiventris</i> , <i>Colaptes punctigula</i> , <i>Phaeomyias murina</i> , <i>Brotogeris sanctithomae</i> , <i>Thamnophilus schistaceus</i> , <i>Turdus ignobilis</i> , <i>Turdus hauxwelli</i> , <i>Myiozetetes cayanensis</i> , <i>Myiozetetes similis</i> , <i>Tyrannus melancholicus</i> , <i>Troglodytes musculus</i>
<i>Entamoeba histolytica</i>	<i>Jacana jacana</i> , <i>Tringa solitaria</i> , <i>Columbina talpacoti</i> , <i>Crotophaga ani</i> , <i>Ammodramus aurifrons</i> , <i>Ramphocelus carbo</i> , <i>Sporophila castaneiventris</i> , <i>Aratinga weddellii</i> , <i>Tyrannus melancholicus</i> .
<i>Entamoeba coli</i>	<i>Jacana jacana</i> , <i>Columbina talpacoti</i> , <i>Crotophaga ani</i> , <i>Dendrocincla fuliginosa</i> , <i>Ammodramus aurifrons</i> .
<i>Echinostoma revolutum</i>	<i>Jacana jacana</i> , <i>Tringa solitaria</i> , <i>Chloroceryle inda</i> , <i>Momotus momota</i> , <i>Crotophaga ani</i> , <i>Sturnella militaris</i> , <i>Leptopogon amaurocephalus</i> , <i>Pheugopedius genibarbis</i> , <i>Pitangus sulphuratus</i> , <i>Turdus ignobilis</i> , <i>Turdus hauxwelli</i> .
<i>Enterobius Vermicularis</i>	<i>Jacana jacana</i> , <i>Tringa solitaria</i> , <i>Columbina talpacoti</i> , <i>Sturnella militaris</i> , <i>Leptopogon amaurocephalus</i> .
<i>Heterakis gallinarum</i>	<i>Turdus ignobilis</i> , <i>Columbina talpacoti</i> , <i>Crotophaga ani</i> , <i>Sturnella militaris</i> , <i>Machaeropterus pyrocephalus</i> , <i>Pipra fasciicauda</i> .

<i>Isoospora</i> sp.	<i>Dendrocygna viduata</i> , <i>Vanellus chilensis</i> , <i>Machaeropterus pyrocephalus</i> , <i>Pipra fasciicauda</i> , <i>Hemitriccus flammulatus</i> , <i>Jacana jacana</i> , <i>Tringa solitaria</i> , <i>Columbina talpacoti</i> , <i>Crotophaga ani</i> , <i>Cacicus cela</i> , <i>Rhynchocyclus olivaceus</i> , <i>Ramphocelus carbo</i> , <i>Sporophila castaneiventris</i> , <i>Tangara episcopus</i> , <i>Catharus swainsoni</i> , <i>Cantorchilus leucotis</i> , <i>Thamnophilus schistaceus</i> , <i>Pheugopedius genibarbis</i> , <i>Turdus ignobilis</i> , <i>Turdus hauxwelli</i> , <i>Tyrannus melancholicus</i> , <i>Troglodytes musculus</i> .
<i>Iodamoeba butschlii</i>	<i>Brotogeris sanctithomae</i>
<i>Microphallus</i> sp.	<i>Jacana jacana</i> , <i>Tringa solitaria</i> , <i>Columbina talpacoti</i> , <i>Turdus ignobilis</i> , <i>Myiozetetes cayanensis</i> , <i>Myiozetetes similis</i> , <i>Tyrannus melancholicus</i> .
<i>Raillietina</i> sp.	<i>Columbina talpacoti</i> , <i>Turdus ignobilis</i> .
<i>Strongyloides avium</i>	<i>Jacana jacana</i> , <i>Momotus momota</i> , <i>Crotophaga ani</i> , <i>Cacicus cela</i> , <i>Pipra fasciicauda</i> , <i>Sporophila castaneiventris</i> , <i>Turdus ignobilis</i> , <i>Myiozetetes similis</i> , <i>Myiozetetes cayanensis</i> .

The present study recorded a considerable diversity (15 species) of endoparasites in the fecal samples of the birds captured in the grounds of the Federal University of Acre (UFAC). These data represent an important advance in the understanding of the diversity and occurrence of endoparasites in the wild bird fauna of Brazil. The high prevalence of parasites recorded in the present study may have important implications for both bird conservation and public health, as well as potential repercussions for the local poultry industry, which reinforces the need for further research.

Acknowledgments: I thank to the Federal University of Acre, the opportunity to study postgraduate, Master's level.

References

- ALLEN, P. C.; FETTERER, R. H. Recent advances in biology of *Eimeria* species and diagnosis and control of infection with these coccidian parasites of poultry. **Reviews Clinical Microbiology**, v. 15, n. 1, p. 58-65, 2002.
- CARLI, G. A. de. **Parasitologia clínica: seleção de métodos e técnicas de laboratório para o diagnóstico das parasitoses humanas**. São Paulo: Atheneu, 2001.
- COMITÊ BRASILEIRO DE REGISTROS ORNITOLÓGICOS. Listas das aves do Brasil. Disponível em: <<http://www.cbro.org.br/Piacentini%20et%20al%202015%20RBO.pdf>>. Acesso em: 25 jul. 2014.
- FREITAS, M. F. L. et al. Parasitos gastrointestinais de aves silvestres em cativeiro em el estado de Pernambuco, Brasil. **Parasitología latinoamericana**, v. 57, n. 1-2, p. 50-54, 2002.
- GIOVANNONI, M.; KUBIK, G. V. L. Fauna parasitológica paranaense. IV. Lista prévia da ocorrência de helmintos em animais domésticos. **Arquivos de Biologia e Tecnologia**, Curitiba, v. 2, n. 1, p. 289-292, 2001.
- GUILHERME, E. Comunidade de aves do Campus e Parque Zoobotânico da Universidade Federal do Acre, Brasil. **Tangara**, v. 1, n. 2 p. 57-73, 2001.
- JACOLEN, G. et al. *Cryptosporidium* sp. em intestino, bursa de Fabricius e traquéia de frangos (*Gallus gallus*). **Ciência rural**, v. 36, n. 2, p. 682-684, 2006.
- LOPES, L. E. **Biologia comparada de *Suiriri affinis* e *Suiriri islerorum* (Aves: Tyrannidae) no cerrado do Brasil central**. 2004. Dissertação (Mestrado) - Universidade de Brasília, Brasília, 2004.
- LUZ, H. R.; FERREIRA, I.; VENTURA, P. E. Estudo da prevalência de ectoparasitos em aves apreendidas no Rio de Janeiro. In: CONGRESSO BRASILEIRO DE ORNITOLOGIA, 12., 2005, Belém - PA. **Anais...** Belém, 2005. p. 160.
- MAPELI, E. B. et al. Infecções naturais por helmintos em perdizes (*Rhynchotus rufescens* Temminck, 1985) de cativeiro, no município de Jaboticabal, estado de São Paulo. **Arquivo Instituto de Biologia**, São Paulo, v. 70, n. 4, p. 415-418, 2003.
- MARIETTO, G. A. G. et al. Prevalência de endoparasitas em amostras fecais de aves silvestres e exóticas examinadas no Laboratório de Ornitopatologia e no Laboratório de Enfermidades Parasitárias da FMVZ-UNESP/Botucatu-SP. **Ciência Animal Brasileira**, v. 10, n. 1, p. 349-354, 2009.
- MARQUES, S. M. T. et al. Parasites of (*Columbia livia*) in urban area of lages, southern Brasil. **Parasitologia Latinoamericana**, v. 62, n. 3-4, p. 183-187, 2007.
- REED, K. D. et al. Birds, migration and emerging zoonoses: west nile virus, lyme disease, influenza a and enteropathogens. **Clinical Medicine and Research**, v. 1, n. 1, p. 5-12, 2003.
- REY, L. **Parasitologia**. Rio de Janeiro: Guanabara Koogan, 2008. p. 856.
- ROSE, M. E. Immunity to *Eimeria* infections. **Veterinary Immunology and Immunopathology**, v. 17, p. 333-343, 1987.
- SILVA, A. S. et al. Parasitismo Por *Isoospora* sp. em *Agapornis fischeri* (Pássaro-do-Amor) criados em cativeiro no Brasil. **Caderno de Pesquisa Série Biologia**, v. 21, n. 1, p. 53-57, 2009.
- SICK, H. **Ornitologia brasileira: uma introdução**. Brasília:

Universidade de Brasília, v. 1, n. 2, p. 827, 1985.

SNAK, A. et al. Análises coproparasitológicas de aves silvestres cativas. **Ciência Animal Brasileira**, v. 15, n. 4, p. 502-507, 2014.

TAYLOR, M. A.; COOP, R. L.; WALL, R. L. **Parasitologia veterinária**. 3. ed. Rio de Janeiro: Guanabara Koogan, 2010.

Recebido em: 20.09.2017

Aceito em: 23.12.2017